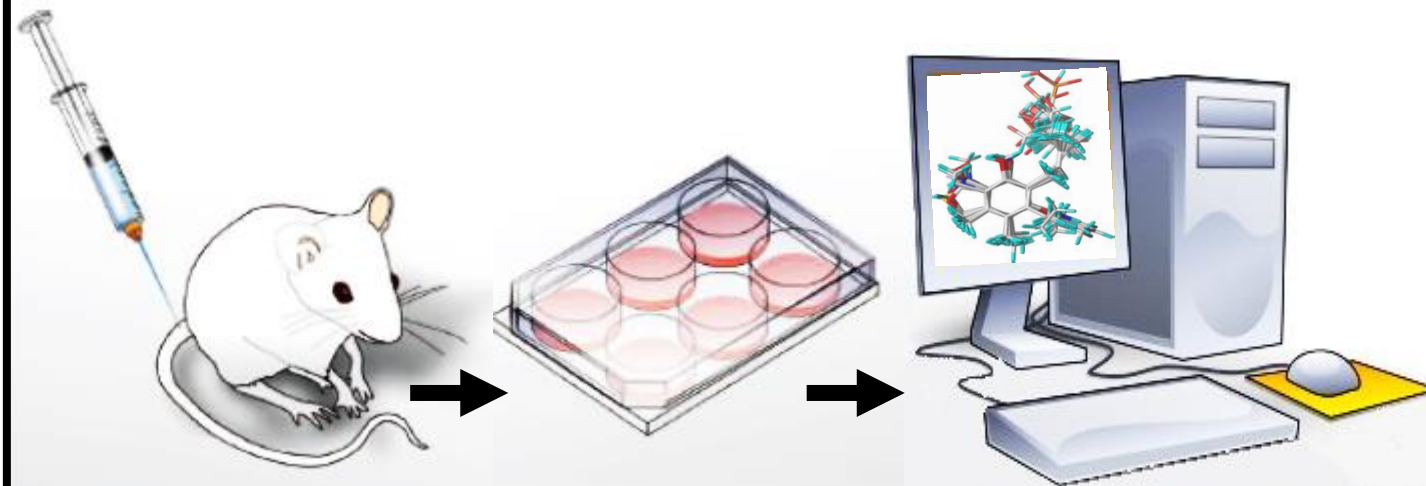


# NanoSAR: Structure-Activity Relationship Model for the Toxicity of **nano** particles



Ceyda OKSEL

Xue Z Wang

# Structure of the lecture

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## ❖ BACKGROUND

- Why are things different at nanoscale ?
- Nanomaterial toxicity
- Computational models for toxicity prediction

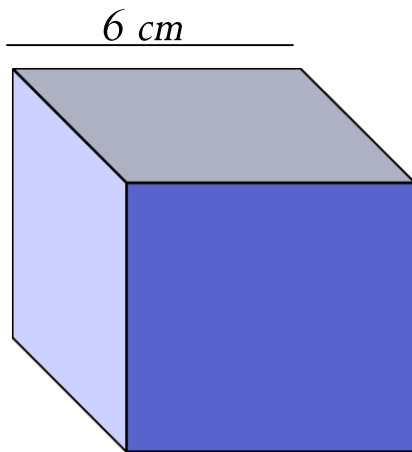
## ❖ COMPUTATIONAL MODELLING OF NANOMATERIAL TOXICITY

- What is (nano)QSAR ?
- 3 Case Studies

## ❖ CONCLUSIONS and FUTURE WORK

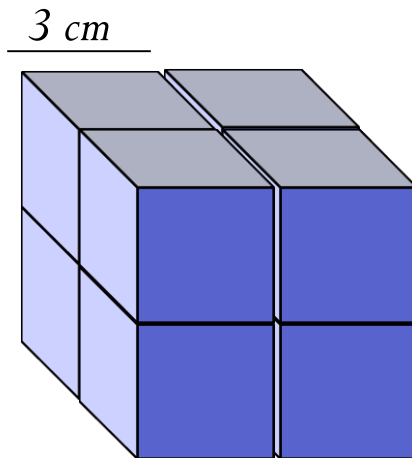
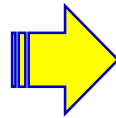
# Why are things different at nanoscale?

Larger surface area



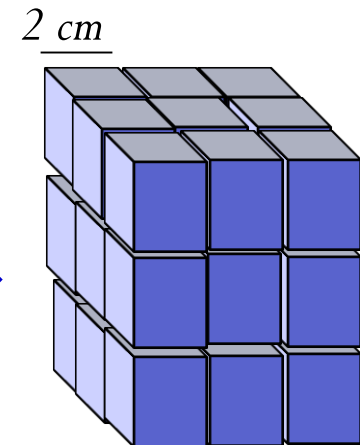
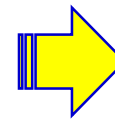
Surface area

$$\begin{aligned} &= (6\text{cm} \times 6\text{cm} \times 6 \text{ faces} \times 1 \text{ cube}) \\ &= 216\text{cm}^2 \end{aligned}$$



Surface area

$$\begin{aligned} &= (3\text{cm} \times 3\text{cm} \times 6 \text{ faces} \times 8 \text{ cubes}) \\ &= 432\text{cm}^2 \end{aligned}$$

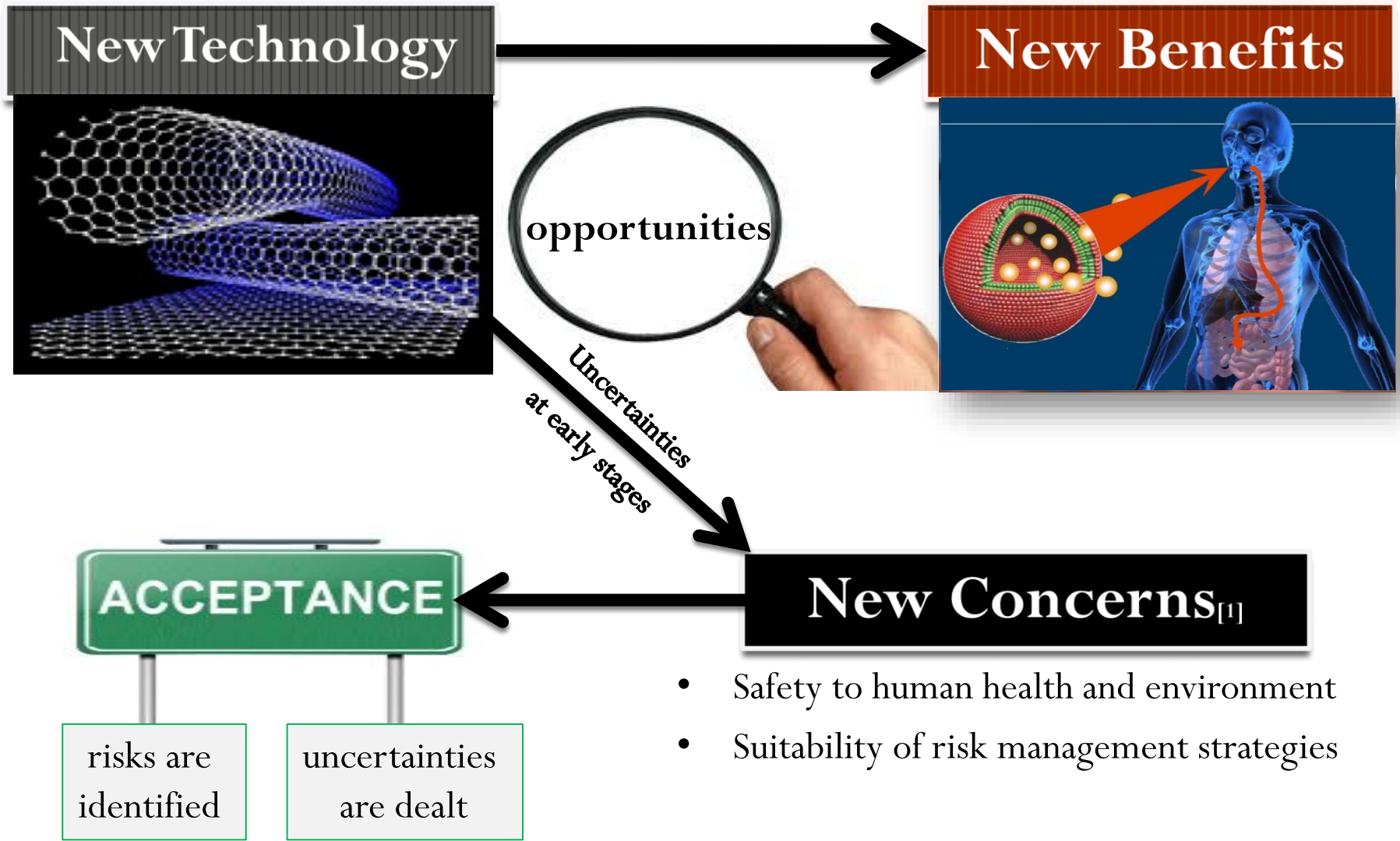


Surface area

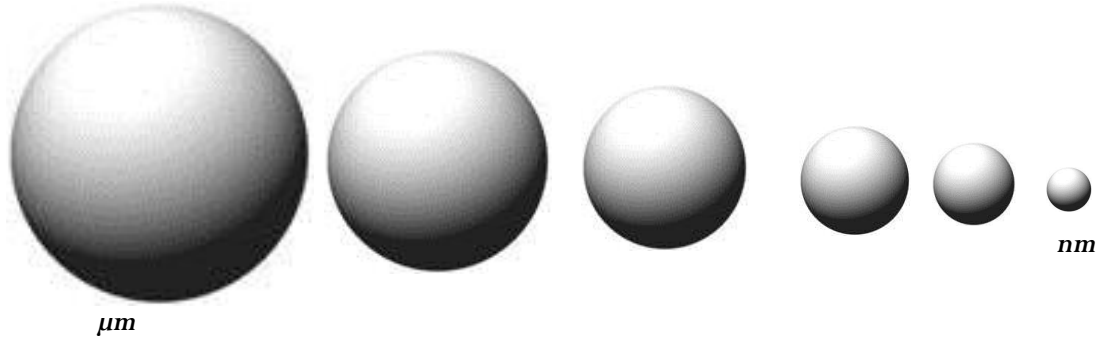
$$\begin{aligned} &= (2\text{cm} \times 2\text{cm} \times 6 \text{ faces} \times 27 \text{ cubes}) \\ &= 648\text{cm}^2 \end{aligned}$$

Quantum effects

# Nanomaterial Toxicity



# Nano Particles, Mega Problems ?



Reduction in particle size

Nano-specific Toxicity

Change in toxicity of NPs

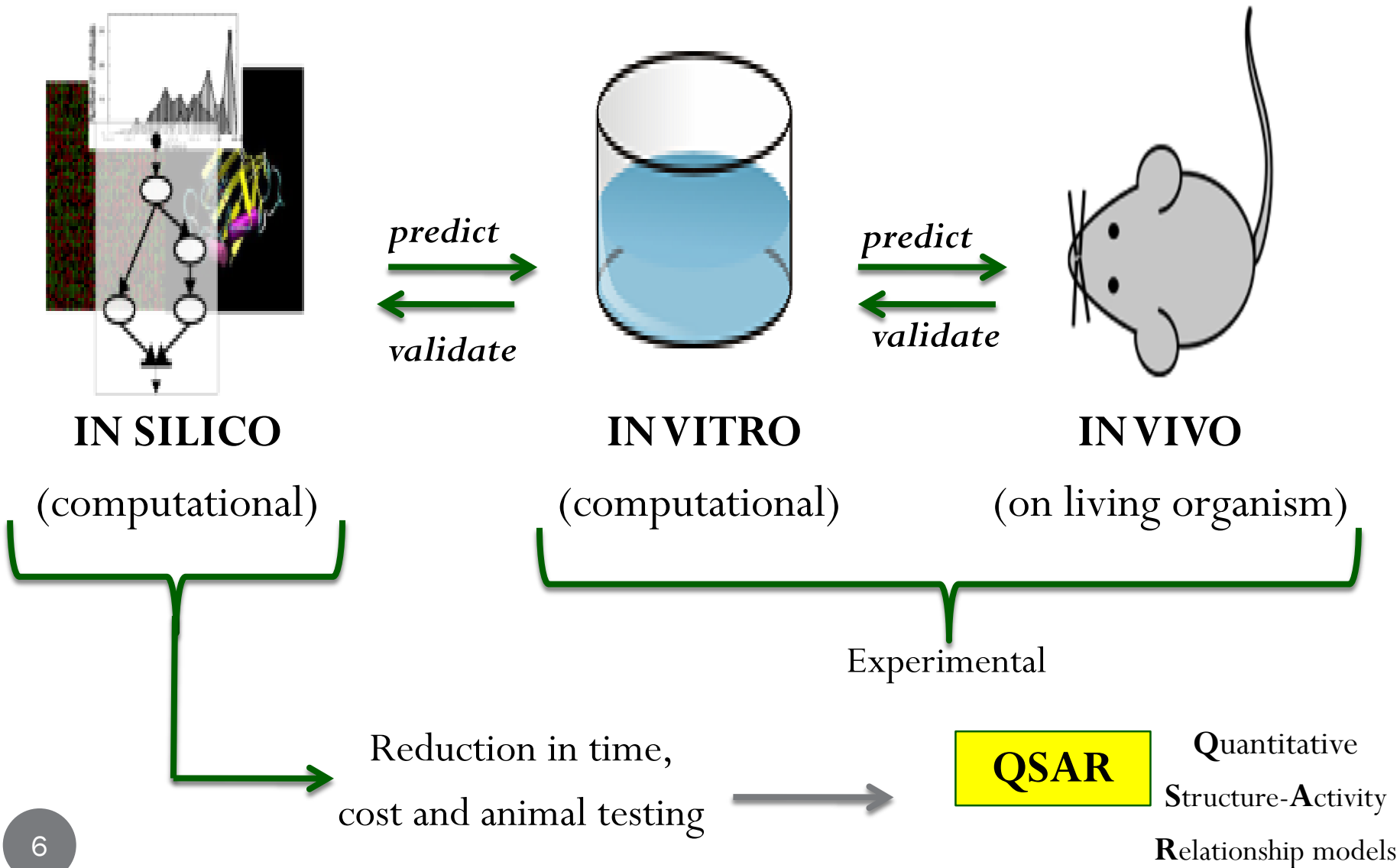
Nano-products

Increase in commercial nanoproducts

Nanosafety Concerns

Increase in nanosafety concerns

# Toxicity Testing



# Why we need computational models?

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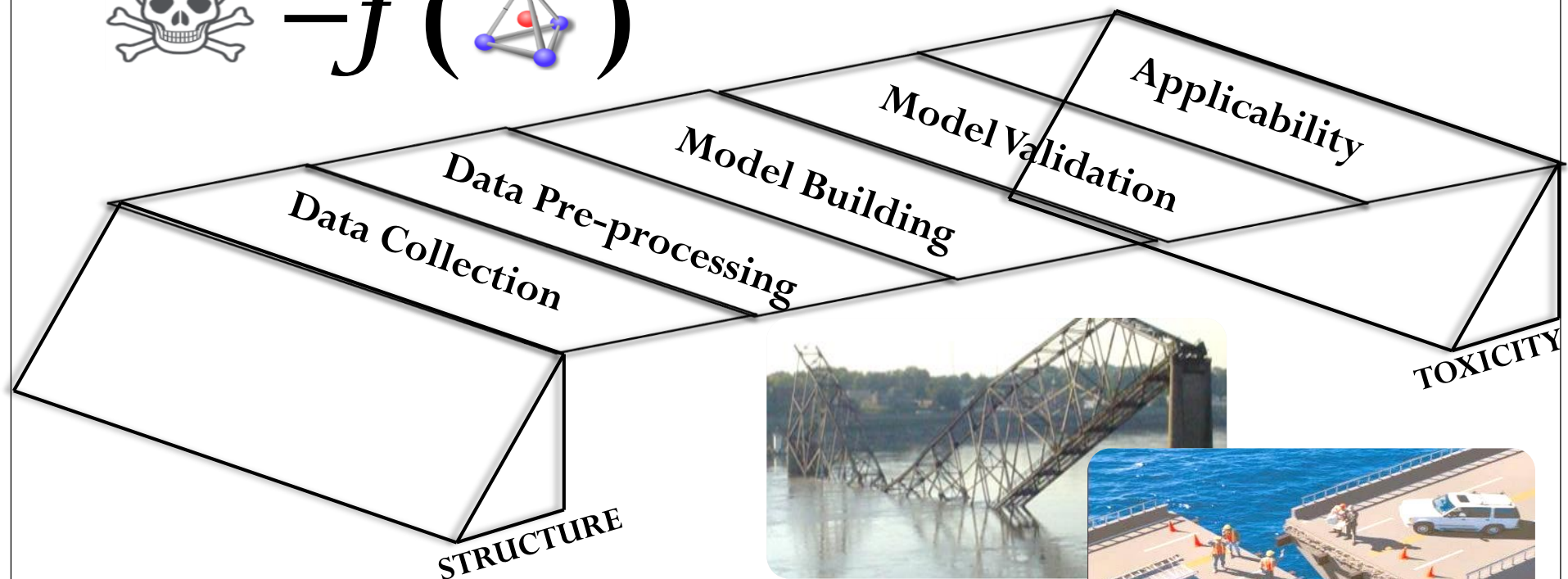


**NEED:** *The European REACH legislation promotes the use of non-animal testing methods*

**AIM: to satisfy this need!!!**

# What is nano-(Q)SAR ?

A (Q)SAR is a statistical model that relates a set of **physicochemical descriptors** of a chemical compound to its **biological activity**.



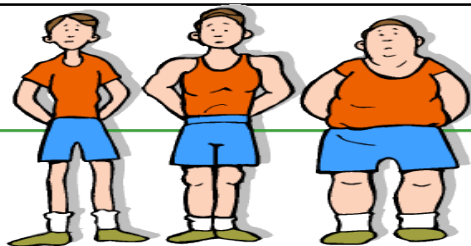
Oksel, C., C.Y. Ma, and X. Z. Wang. "Current situation on the availability of nanostructure–biological activity data." *SAR and QSAR in Environmental Research* ahead-of-print (2015): 1-16.

Oksel, C., C.Y. Ma, J. J. Liu, T. Wilkins, X. Z. Wang, (2015) (Q)SAR modelling of nanomaterial toxicity: A critical review, *Particuology*, 10.1016/j.partic.2014.12.001



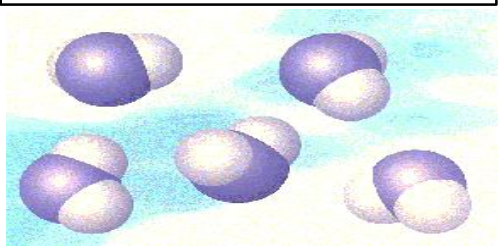
# Descriptors

## DESCRIBING A PERSON



- ✓ Height
- ✓ Weight
- ✓ Attractiveness
- ✓ ...
- ✓ Eye
- ✓ Hair
- ✓ Build
- ✓ ...

## DESCRIBING A MOLECULE



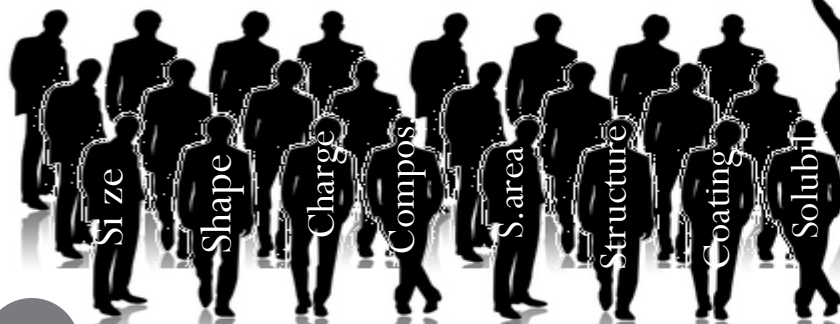
- ✓ Molar mass
- ✓ Density
- ✓ Conductivity
- ✓ ...
- ✓ Atomic prop.
- ✓ Bonds
- ✓ Chirality
- ✓ ...

## DESCRIBING A NANOPARTICLE



- ✓ Size
- ✓ Shape
- ✓ Composition
- ✓ ...
- ✓ Coating
- ✓ Charge
- ✓ Reactivity
- ✓ ...

## Experimental Descriptors

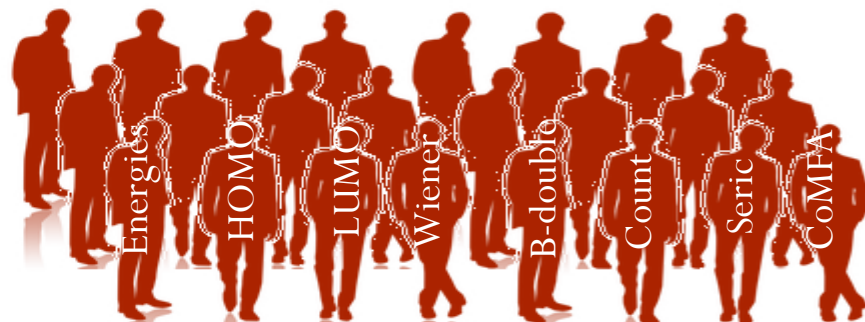


## Descriptor Selection

Feature selection algorithms



## Theoretical Descriptors



# Tree Induction From Genetic Programming

## **GPTree**: “in-house” software

Genetic Algorithms

**explore  
solution space**

- Starts at random points
- Recombining (i.e., crossover)
- Optionally changing (i.e., mutation)

**Genetic  
Algorithm**

- (1) Randomly generate a pre-specified number of solutions, encoded as fixed size vectors.**
- (2) Either form a new generation or replace individuals in the population by**
  - 2a. Selecting parents using the fitness function.**
  - 2b. Crossover the parents to form one or more offspring.**
  - 2c. Optionally mutate part of the solution.**
- (3) Continue with Step 2 until a pre-specified number of generations or children have been grown, or until a good solution is found.**

# Tree Induction From Genetic Programming

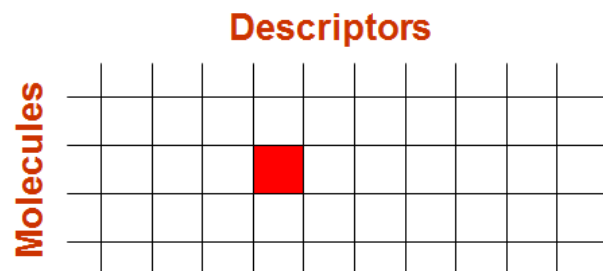
## GP Tree: Methodology

- DeLisle, R. K. and Dixon, S. L. (2004) Induction of Decision Trees via Evolutionary Programming *Journal of Chemical Information and Computer Sciences*, 44, 862-870.- **evolutionary programming of trees**

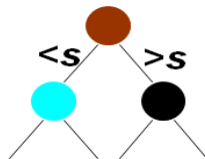
### 1. Divide data into training and test sets

### 2. Generate the 1<sup>st</sup> population of trees

- randomly choosing a row (i.e. a compound), and column (i.e. descriptor)



- Using the value of the slot,  $s$ , to split, left child takes those data points with selected attribute values  $\leq s$ , whilst the right child takes those  $> s$ .



# Tree Induction From Genetic Programming

## **GP**Tree: Methodology

- If a child will not cover enough rows (e.g. 10% of the training rows), another combination is tried.
- A child node becomes a leaf node if pure/near pure, whilst the other nodes grow children.
- When all nodes either have two children or are leaf nodes, the tree is fully grown and added to the first generation.
- A leaf node is assigned to a class label corresponding to the majority class of points partitioned there.

### **3. Crossover and Mutation**

# Tree Induction From Genetic Programming

## The key parameters

<b>y COL</b>	Column no containing the class of the data set.
<b>n Gen</b>	No of generations required
<b>n Trees</b>	No of trees required in each generation
<b>No. in tournament</b>	No of trees in the tournament to sort out the best for crossover operation
<b>Winn. Inc.</b>	Winners included (The N best trees are placed directly into the next generation, This was to allow ELITISM)
<b>L.I.I.A.T</b>	Low increase in accuracy tolerance (It forces a mutation for every tree if no improvement in the best accuracy has been seen for this many generations.)
<b>Mutation</b>	% age of mutation
<b>C in L.N</b>	Minimum no of cases in a leaf node

# Case Study 1: Dataset

## Compounds

75 Compounds

## Toxicity Data (4 classes)

Concentration lethal to 50% of the population, LC50,  
1/Log(LC50), of *vibrio fischeri*, a bioluminescent bacterium

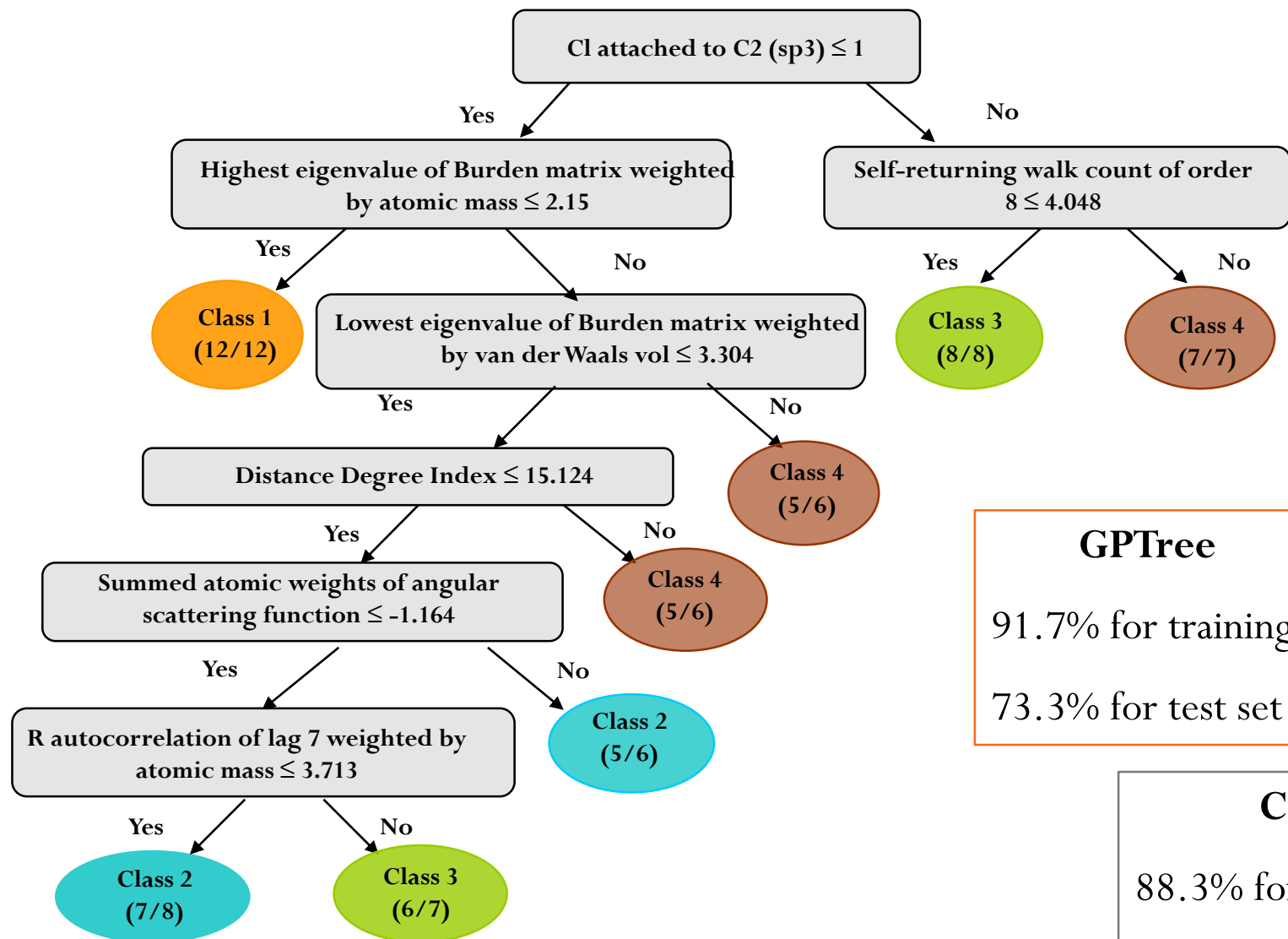
## Descriptors

1069 molecular descriptors calculated by DRAGON

## Parameters

y COL	1070
n Gen	60
n Trees	600
No. in tournament	16
Winn. Inc.	0
L.I.I.A.T	5
Mutation	66.7%
C in L.N	2

# Case Study 1: Results



## GPTree

91.7% for training

73.3% for test set

## C5.0

88.3% for training

60.0 % for test set

# Case Study 2: Dataset

## Compounds

105 nanoparticles with different surface-modifying molecules

## Toxicity Data

Cellular uptake in pancreatic cancer cell lines

## Threshold value

Cellular uptake values: 170-27 542 nanoparticles per cell

Threshold value: 10 000 nanoparticles per cell

18 nanoparticles with significant cellular uptake (CLASS 2)

87 nanoparticles with poor cellular uptake (CLASS 1)

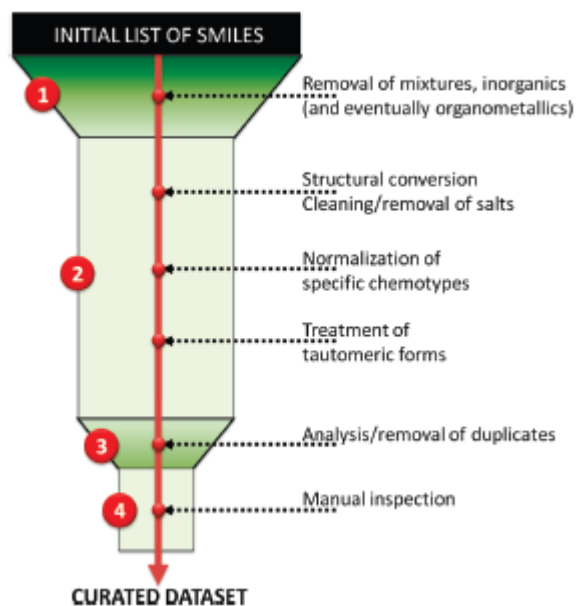


# Case Study 2: Dataset

## Descriptors

Nanoparticles  $\longrightarrow$  Different surface-modifying molecules  $\longrightarrow$  Conventional descriptors

Same core

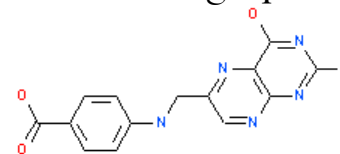


Fourches et al. (2010)

- **Data cleaning**
- **Structural Conversion**

SMILES strings  $\longrightarrow$  2D molecular graphs

[C]=NC(=C(N=1)C1O)N=C(N=1)N)CNC(=CC=C(C1)C(O)=O)C



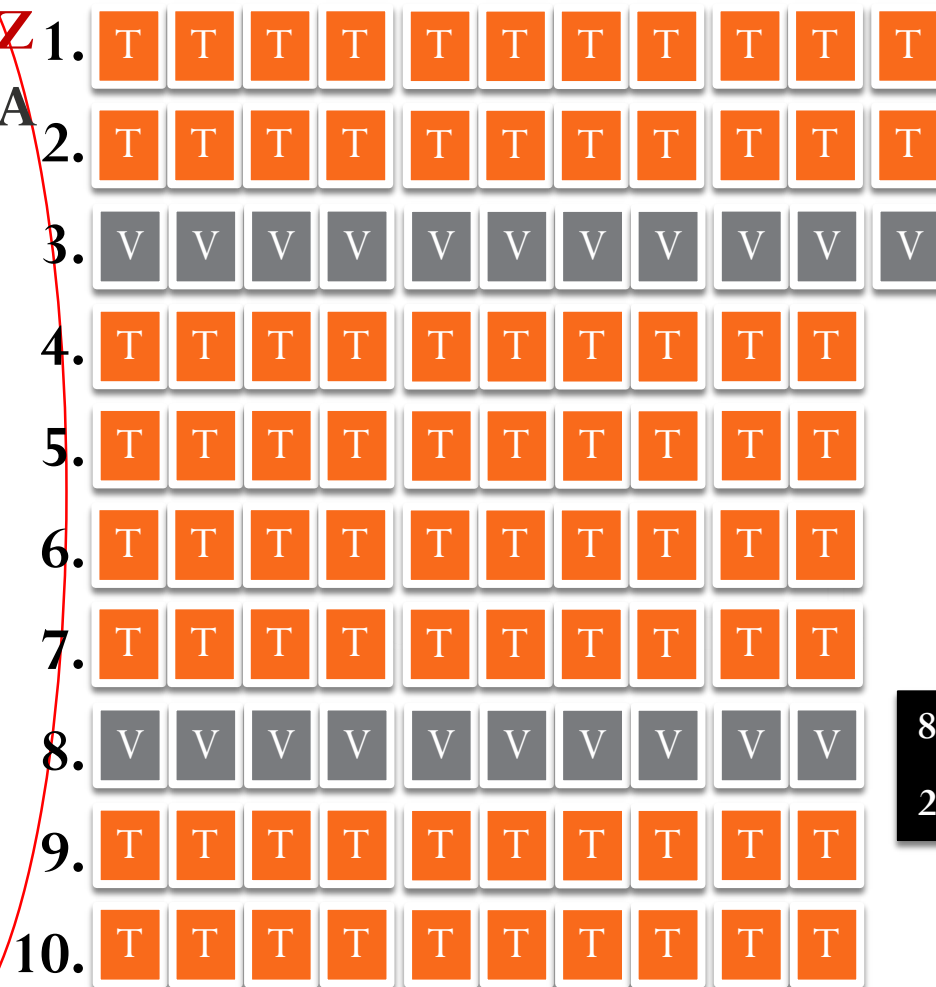
- **Manual inspection**  
4 structure unmatched-excluded
- **Descriptor Calculation**  
690 Dragon Descriptors
- **Descriptor Cleaning**  
389 Dragon descriptors retained

# Case Study 2: Data Pre-processing

## Data splitting

No	Smiles Notation	Cellular uptake
35	<chem>O=C1Nc2cccc2C(=O)O1</chem>	27542
71	<chem>CCCCCCCCCCCCCN</chem>	18621
98	<chem>O=C1CCC(=O)O1</chem>	17378
17	<chem>Clc1ccc2NC(=O)OC(=O)c2c1</chem>	15136
1	<chem>FC(F)(F)C(=O)OC(=O)C(F)(F)F</chem>	14791
21	<chem>Clc1cc2C(=O)OC(=O)c2cc1Cl</chem>	1318
4	<chem>CC1(C)CC(=O)OC1=O</chem>	12882
15	<chem>O=C1OC(=O)c2ccc(cc3cccc1c23)N(=O)=O</chem>	12882
105	<chem>OC(=O)CN(CCN1CC(=O)OC(=O)C1)CCN1CC(=O)OC(=O)C1</chem>	12589
3	<chem>FC(F)(F)C(F)(F)C(=O)OC(=O)C(F)(F)C(F)(F)F</chem>	12023
33	<chem>O=C1CCCC(=O)O1</chem>	11749
62	<chem>CC(C)(C)N</chem>	11749
48	<chem>O=C1CC2(CCCC2)CC(=O)O1</chem>	11482
64	<chem>CCCCCCCCCCCCCN</chem>	11482
99	<chem>CC(=O)OC(C)=O</chem>	11220
100	<chem>C=C1CC(=O)OC1=O</chem>	10965
47	<chem>CCCCCCCCC(=O)OC(=O)CCCCCCCC</chem>	10715
103	<chem>OC(=O)CC1CC(=O)OC1=O</chem>	10715
50	<chem>O=C1CCC(C(=O)O1)c1ccccc1</chem>	10471
101	<chem>O=C1COCC(=O)O1</chem>	9772
5	<chem>O=C1OC(=O)C=C1</chem>	9550
30	<chem>Cc1ccc2C(=O)OC(=O)c2c1</chem>	9550
16	<chem>Oc1ccccc2C(=O)OC(=O)c1c2</chem>	9333
69	<chem>CCCCCCCCCCCCCCCCCN</chem>	9333
49	<chem>O=C1OC(=O)c2ccccc3cccc1c23</chem>	9120
2	<chem>FC(F)(Cl)C(=O)OC(=O)C(F)(F)Cl</chem>	8913
68	<chem>CCCCC(CC)CN</chem>	8913
53	<chem>CC1(C)CCC(=O)OC1=O</chem>	8710
26	<chem>O=C1OC(=O)c2ccc(cc3cccc1c23)N(=O)=O</chem>	8511
34	<chem>O=C1CN(CCN2CC(=O)OC(=O)C2)CC(=O)O1</chem>	8511
41	<chem>O=C1OC(=O)C2CCCC12</chem>	8511
37	<chem>CC1CC(=O)OC(=O)C1</chem>	8128
39	<chem>CC(=O)OC1C(OC(C)=O)C(=O)OC1=O</chem>	8128
104	<chem>Fe1ccc(F)c2C(=O)OC(=O)c1c2</chem>	8128
52	<chem>Clc1ccc(Cl)c2C(=O)OC(=O)c1c2</chem>	7943
102	<chem>O=C1OC(=O)c2ccccc12</chem>	7943
24	<chem>O=C1OC(=O)C2CC=CCC12</chem>	7762
18	<chem>O=C1OS(=O)(=O)c2ccccc12</chem>	7586
58	<chem>CC(C)(C)N</chem>	7244
55	<chem>CC(C)CC(C)N</chem>	7079
19	<chem>ClC1=C(C)C(=O)OC1=O</chem>	6918
14	<chem>Fe1c(F)c(F)c2C(=O)OC(=O)c2c1F</chem>	6761
51	<chem>Clc1c(Cl)c(Cl)c2C(=O)OC(=O)c2c1Cl</chem>	6761
60	<chem>CC(C)CCN</chem>	6761
22	<chem>O=C1OC(=O)C2C3OC(C=C3)C12</chem>	6607
28	<chem>CCCCCCCCCCCC(=O)OC(=O)CCCCCCCCCCC</chem>	6607
61	<chem>CC(N)CC</chem>	6457
40	<chem>Brc1c(Br)c(Br)c2C(=O)OC(=O)c2c1Br</chem>	6310

## Pattern of splitting



80% training  
20% validation

# Case Study 2: GP Tree settings

## The key parameters

```
1 EPTREE Train.txt Test.txt 390 60 600 16 0 5 1 2 2
```

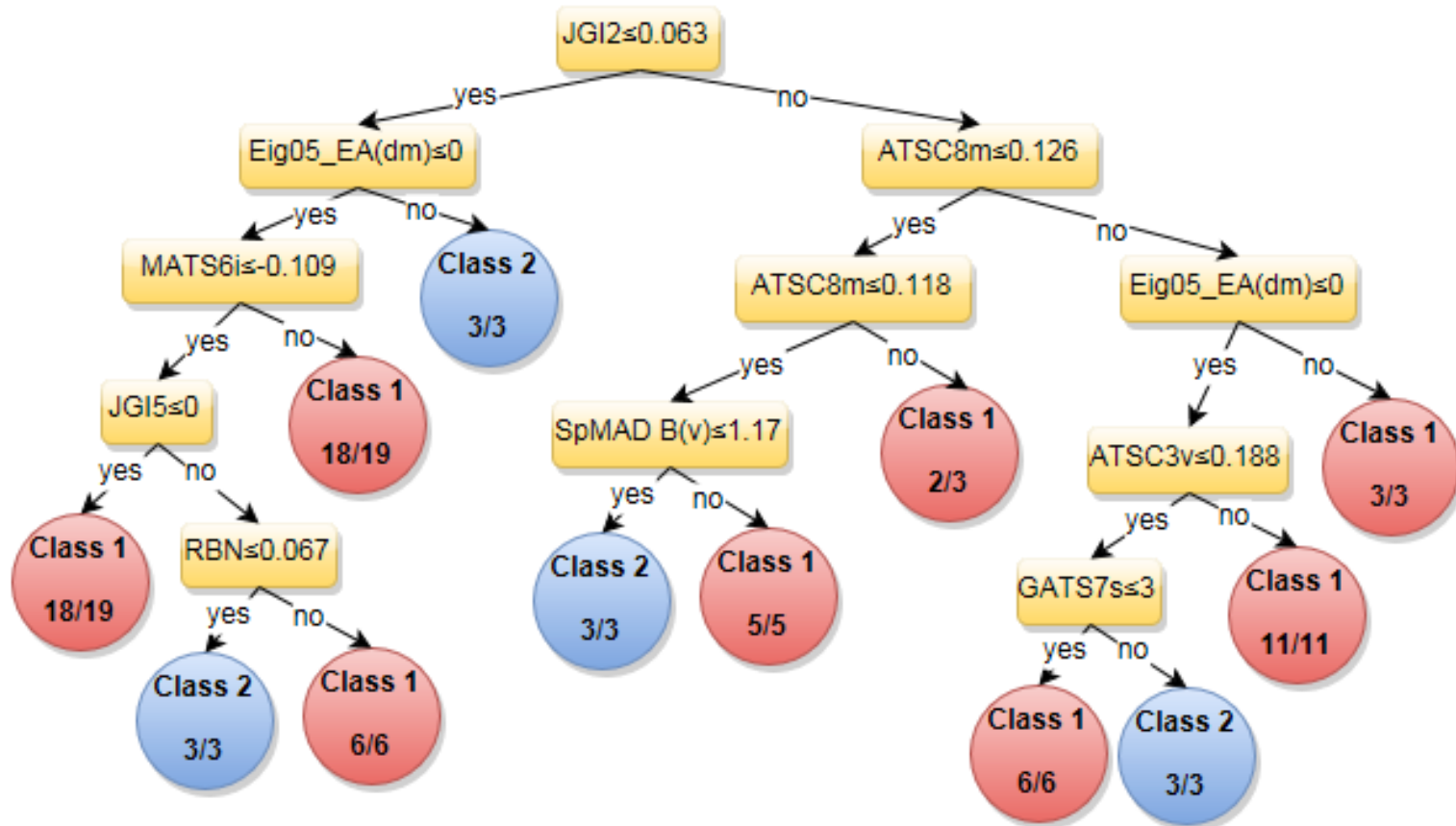
Column no containing the class of the data set	390
No of generations required	60
No of trees in each generation required	600
No of trees in the tournament	16
Winners included	0
Low increase in accuracy tolerance	5
% age of mutation	50%
Minimum no of cases in a leaf node	2

# Case Study: Results

## GPTree Results

```
Best.txt - Notepad
File Edit Format View Help
Gen 22 Tree 38
xover: 1 tree 26 rtree 309 idL 20, idR 21
[0] col 171 val 0.063000 (from row 27)
Left
  Parent -1 Left 9 Right 1
  Train ClassFreq: [1: 69], [2: 15],
  Test ClassFreq: [1: 17], [2: 4],
[1] col 108 val 0.126000 (from row 7)
Right
  Parent 0 Left 4 Right 2
  Train ClassFreq: [1: 27], [2: 7],
  Test ClassFreq: [1: 9], [2: 1],
[2] col 230 val 0.000000 (from row 40)
Right
  Parent 1 Left 18 Right 3
  Train ClassFreq: [1: 20], [2: 3],
  Test ClassFreq: [1: 9], [2: 1],
[3] Leaf node
Right
  Parent 2 Left -1 Right -1
  Train ClassFreq: [1: 3],
  Test ClassFreq: [none covered]
  Train rows covered:
  8, 25, 33,
  Test rows covered:
[4] col 108 val 0.118000 (from row 24)
Left
  Parent 1 Left 5 Right 8
  Train ClassFreq: [1: 7], [2: 4],
  Test ClassFreq: [none covered]
[5] col 92 val 1.170000 (from row 7)
Left
  Parent 4 Left 7 Right 6
  Train ClassFreq: [1: 5], [2: 3],
  Test ClassFreq: [none covered]
[6] Leaf node
Right
  Parent 5 Left -1 Right -1
  Train ClassFreq: [1: 5],
  Test ClassFreq: [none covered]
  Train rows covered:
  3, 42, 46, 54, 59,
  Test rows covered:
[7] Leaf node
Left
  Parent 5 Left -1 Right -1
  Train ClassFreq: [2: 3],
  Test ClassFreq: [none covered]
  Train rows covered:
  24, 76, 77,
  Test rows covered:
[8] Leaf node
Right
  Parent 4 Left -1 Right -1
  Train ClassFreq: [1: 2], [2: 1],
  Test ClassFreq: [none covered]
  Train rows covered:
  4, 7, 78,
  Test rows covered:
[9] col 230 val 0.000000 (from row 40)
Left
  Parent 0 Left 11 Right 10
  Train ClassFreq: [1: 4], [2: 8],
  Test ClassFreq: [1: 4], [2: 8],
  Train rows covered:
  5, 14, 28, 31, 79, 80,
  Test rows covered:
  5, 7, 8, 9,
[22] Leaf node
Right
  Parent 20 Left -1 Right -1
  Train ClassFreq: [2: 3],
  Test ClassFreq: [1: 1], [2: 1],
  Train rows covered:
  15, 35, 81,
  Test rows covered:
  0, 4,
** Total covered 84, Leaf nodes 12 Accuracy 96.428571
Test Total covered 21, Accuracy 80.952381
```

# Case Study: Results



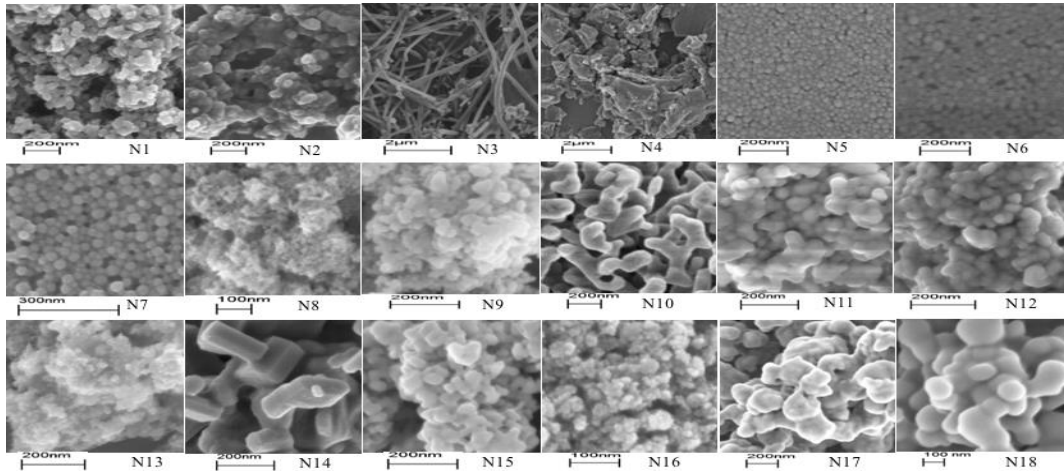
**Training accuracy: 96%**  
**Test accuracy: 81%**

**9 descriptors out of 389**

# Case Study: Results

DRAGON descriptor	Description	Block
JGI2	mean topological charge index of order 2	2D autocorrelations
JGI5	mean topological charge index of order 5	2D autocorrelations
ATSC8m	Centred Broto-Moreau autocorrelation of lag 8 weighted by mass	2D autocorrelations
ATSC3v	Centred Broto-Moreau autocorrelation of lag 3 weighted by van der Waals volume	2D autocorrelations
MATs6i	Moran autocorrelation of lag 6 weighted by ionization potential	2D autocorrelations
GATS7s	Geary autocorrelation of lag 7 weighted by I-state	2D autocorrelations
Eig05_EA(dm)	eigenvalue n. 5 from edge adjacency mat. weighted by dipole moment	Edge adjacency indices
SpMAD B(v)	spectral mean absolute deviation from Burden matrix weighted by van der Waals volume	2D matrix-based descriptors
RBN	number of rotatable bonds	Constitutional indices

# Case Study 3: Data Collection



Carbon Black <b>N1</b>	Aluminum Oxide <b>N10</b>
Diesel Exhaust <b>N2</b>	Cerium Oxide <b>N11</b>
Japanese Nanotubes <b>N3</b>	Nickel Oxide <b>N12</b>
Fullerene <b>N4</b>	Silicon Oxide <b>N13</b>
Polystyrene Latex Beads <b>N5</b>	Zinc Oxide <b>N14</b>
Polystyrene Latex Beads <b>N6</b>	Titanium Dioxide Rutile <b>N15</b>
Polystyrene Latex Beads <b>N7</b>	Titanium Dioxide Anatase <b>N16</b>
Aluminum Oxide <b>N8</b>	Silver <b>N17</b>
Aluminum Oxide <b>N9</b>	Silver <b>N18</b>

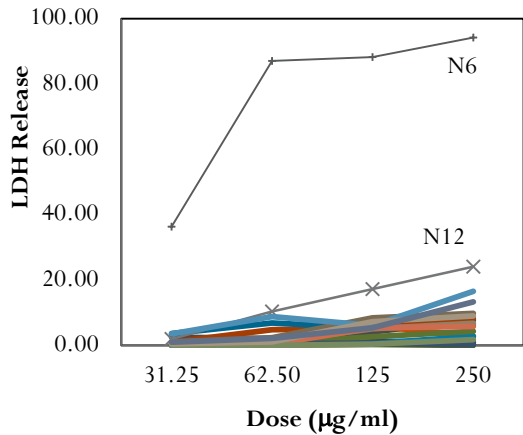
## Characterization

- **Particle size and size distribution** were analysed using a Malvern MasterSizer 2000
- **Particle shape** was analysed using LEO 1530 Scanning Electron Microscope (SEM) or Philips CM20 Transmission Electron Microscope (TEM)
- **Surface area and porosity** were measured using TriStar 3000 BET
- **The free radical activities** were measured by EPR
- **Particle reactivity in solution**, the dithiothreitol (DTT) consumption
- **Metal Content** was measured
- **Charge:** z potential was measured using Malvern Instrument's Zetasizer Nano instrument

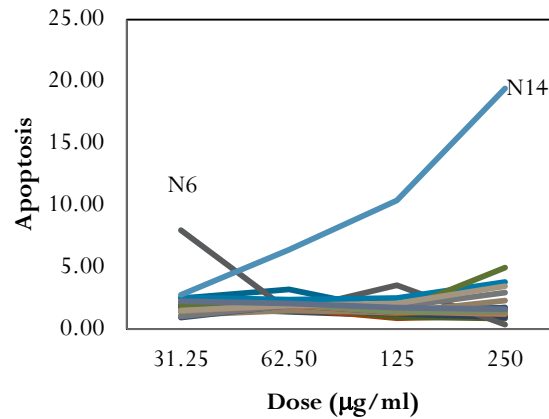
# Case Study 3: Data Collection

## Toxicological Evaluation

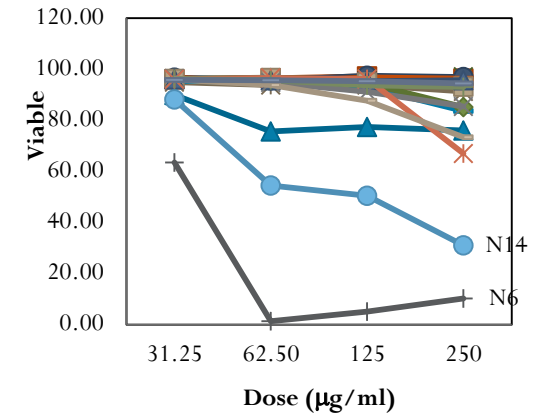
### LDH Release



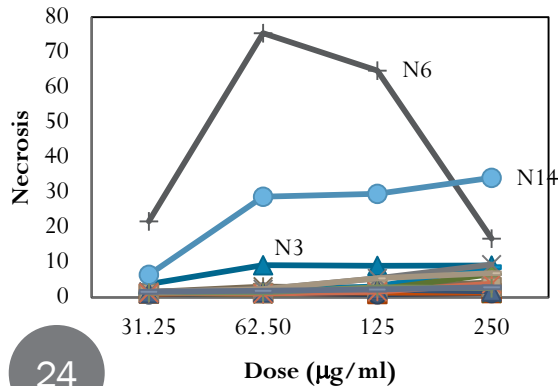
### Apoptosis



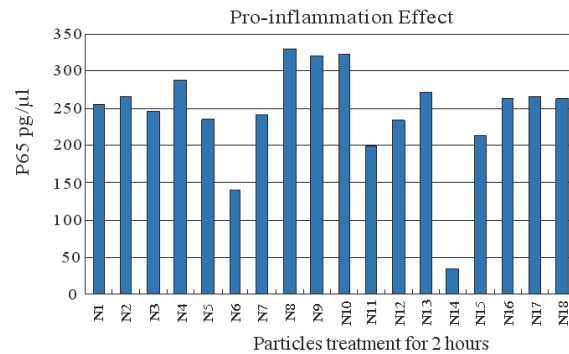
### Viability



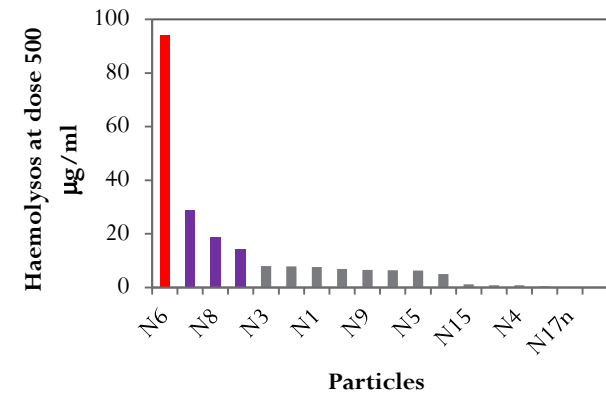
### Necrosis



### Pro-inflammation effects



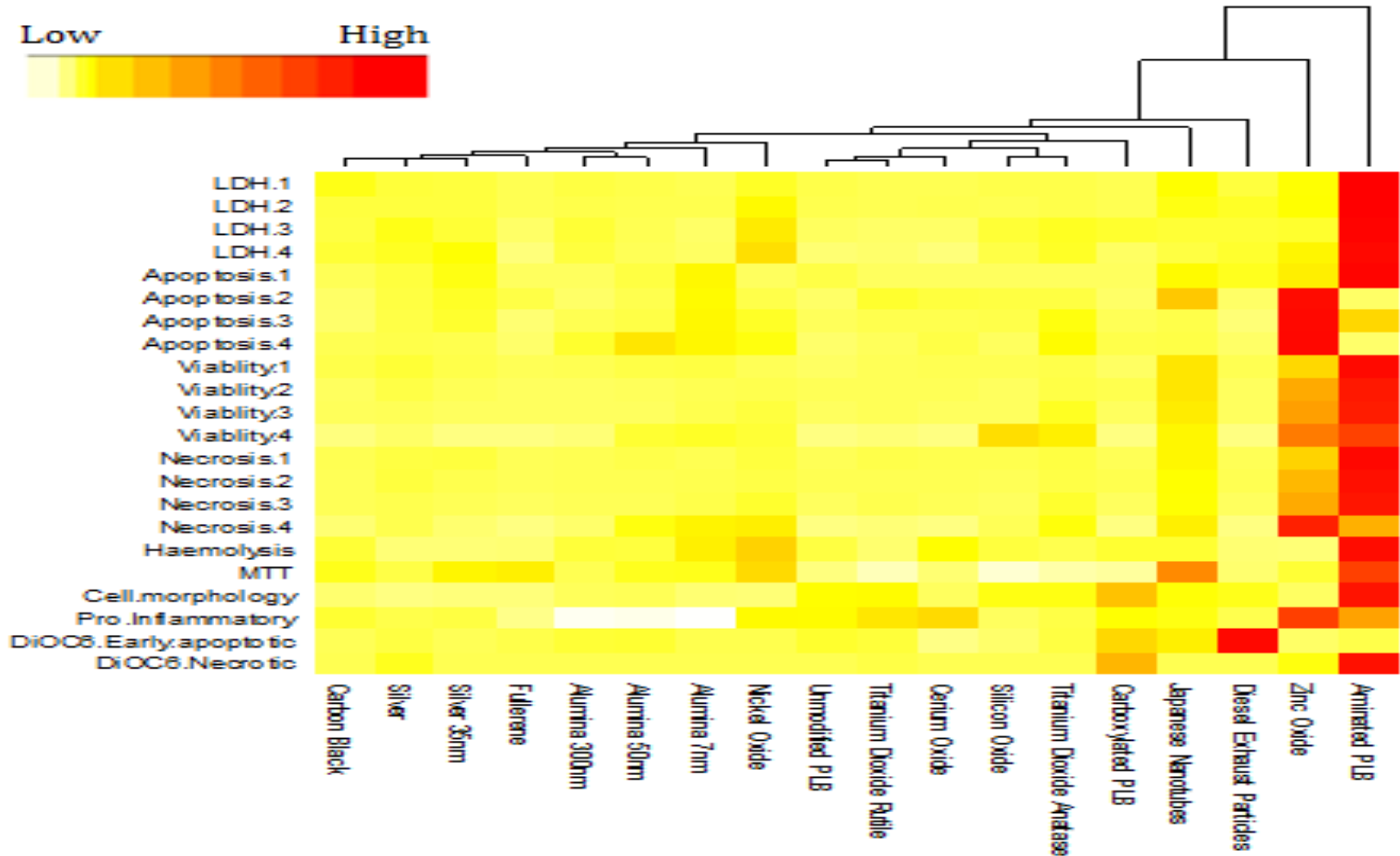
### Haemolysis





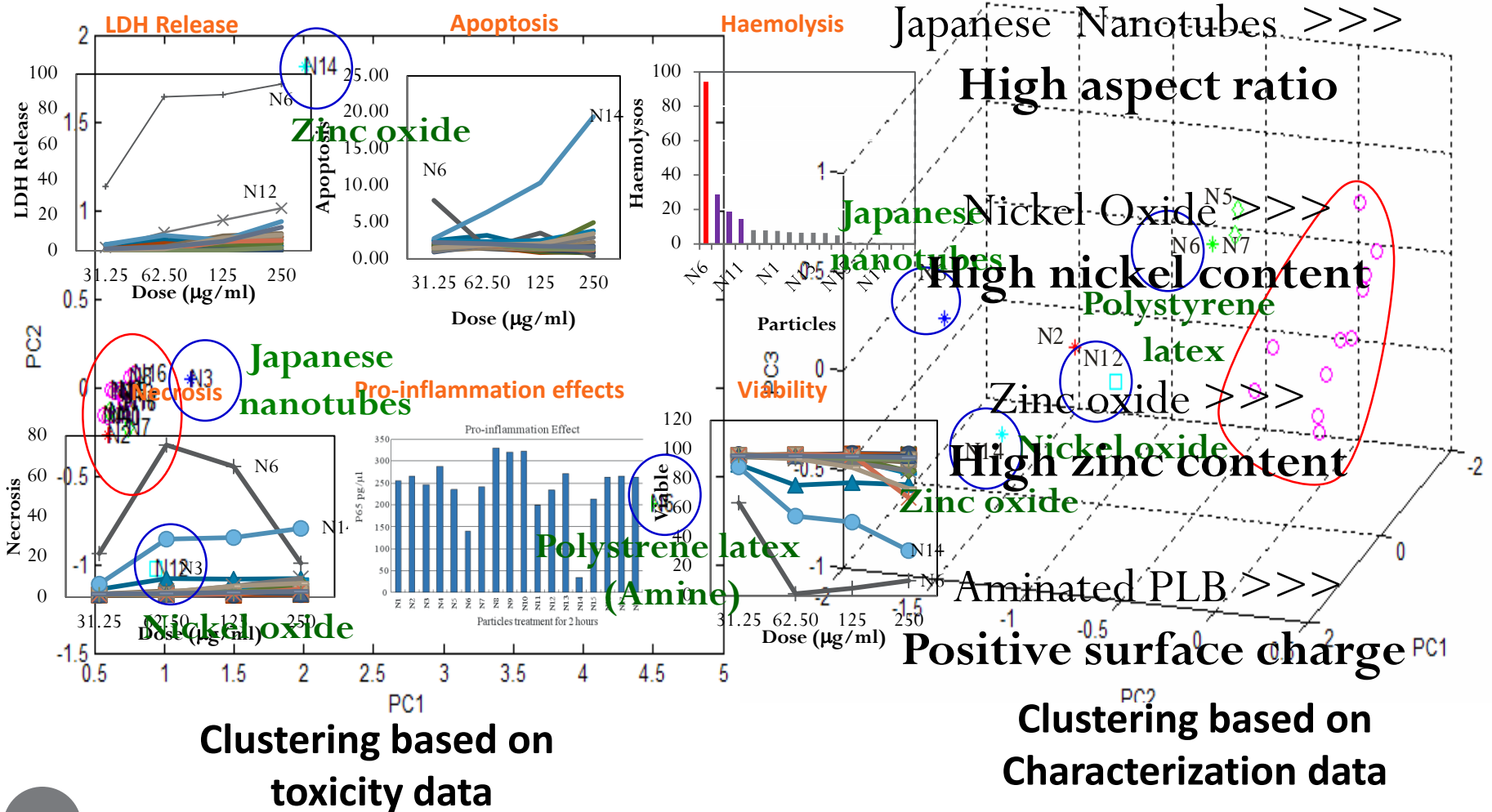
# Case Study 3: Data Visualization

## Multidimensional data visualization: Heat maps with hierarchical clustering



# Case Study3: Model Development

## Clustering/Grouping based on Principal Component Analysis



# Conclusions

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- In LEEDS, we have developed a decision tree software which can be successfully employed for nano-(Q)SAR investigations
- (Q)SAR tools are useful for identifying the properties that influence the toxicity
- Many potential profits:
  - An alternative, fast and cheap way of hazard assessment
  - Risk Reduction
  - Safety-by-design

# Future Work

No	Dataset	Nanomaterials	Toxicity Endpoint	Characterization
1	<a href="#">Wang et al. (2014)</a>	18 NMs (carbon-based and metal oxides)	LDH release, apoptosis, pro-inflammatory effects, haemolysis, MTT, DiOC6, cell morphology assay	size, surface area, morphology, metal content, reactivity, free radical generation and zeta potential
2	<a href="#">Shaw et al. (2008)</a>	50 NMs with diverse core structures	ATP content, reducing equivalents, apoptosis, mitochondrial membrane potential	core composition, coating type, surface modification, size, relaxivities and zeta potential
3	NANOMMUNE project	18 NMs	In vitro assays	core, coating, 2 sizes and zeta potential
4	<a href="#">Puzyn et al. (2011)</a>	17 metal oxide NMs	Cytotoxicity (EC50)	12 different quantum-mechanical descriptors
5	MARINA project	9 NMs	In vitro assays	experimental descriptors
6	<a href="#">Weissleder et al. (2005)</a>	109 NMs with the same core but different surface modifiers	Cellular uptake	theoretical descriptors
7	B.Yan (private communication)	80 surface-modified MWCNTs	Protein binding activities, cell viability, nitrogen oxide generation	theoretical descriptors
8	<a href="#">Liu et al. (2011)</a>	9 metal oxide NMs	Cytotoxicity (PI uptake)	a set of 10 descriptors
9	<a href="#">Sayes and Ivanov (2010)</a>	42 NMs with two cores (differing in concentrations)	Cellular membrane damage (LDH release)	primary particle size, size in water and buffered solutions, concentration and zeta potential
10	ENPRA project	10 NMs	In vitro/in vivo assays	size, dustiness, surface area and impurities
11	<a href="#">Gajewicz et al. (2014)</a>	18NMs	Cellular viability (LC50)	18 quantum mechanical descriptors, 11 image descriptors, 3 experimental descriptors



~~NANO-FEAR~~

SUSTAINABILITY of NANOTECHNOLOGY

Thank you !